

**invivo**<sup>®</sup>

# Oral EcologiX<sup>™</sup>

## Oral Health & Microbiome Profile

Phylo Bioscience Laboratory

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### INTERPRETIVE GUIDE

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# Oral EcologiX™

## INTRODUCTION

Due to recent advancements in culture-independent techniques, it is now possible to measure the composition of the human microbiota. The oral cavity is a complex ecosystem, comprising several habitats including the teeth, gums, tongue and tonsils, all colonised by bacteria<sup>1</sup>. The oral microbiota is comprised of approximately 600 taxa at the species level, with different groups and subsets inhabiting different niches.

The microbiota of the oral cavity exists as a complex biofilm that remains stable despite environmental changes. However, dysbiosis, in form of infection, injury, dietary changes and risk-associated factors (e.g. smoking) may disrupt the biofilm community, favouring colonisation and invasion of pathogens. Disruption of the biofilm community to a pathogenic profile, induces host immune responses, chronic inflammation and ultimately development of local and systemic diseases. However, much of this damage is reversible if pathogenic communities are removed, and homeostasis is restored.

To this end, Phylbioscience have developed the Oral EcologiX™ oral health and microbiome profile, a ground breaking tool for analysis of oral microbiota composition and host immune responses. Using innovative microbial culture-independent technologies, including quantitative real-time PCR (qRT-PCR) and enzyme-linked immunosorbent assay (ELISA), the profile provides an accurate, reliable and quantifiable measurement of microbiota abundance and host inflammatory markers.

### **For microbiota composition analysis, the technology detects:**

- Abundance of commensal bacteria
- Abundance of caries-associated bacteria
- Abundance of periodontitis-associated bacteria
- Presence of viral pathogen herpes simplex virus 1 and 2 (HSV-1)
- Presence of human papillomavirus-16 genotype (HPV-16)

### **For host biomarker analysis, the technology detects:**

- IL-1β (marker for inflammation)

### **Dependent on microbiota composition and host biomarkers, the Oral EcologiX™ profile will report three different states:**

- Healthy oral microbiome (homeostasis of host and microbiome)
- Oral dysbiosis detected (imbalance of the oral microbiota and/or host immune response)
- Oral pathogen detected (HSV-1, HPV-16)

## GLOSSARY

Term	Description	References
Commensal	Microorganism (i.e. bacteria, fungi) that lives in symbiosis with the host when residing within its specific environment	1
Pathogen	Microorganism (e.g. bacteria, fungi, virus) that may cause disease	1
Homeostasis	Ability to maintain internal stability in an organism despite environmental changes	2
Dysbiosis	Imbalance or disturbance in the human microbiota	3
Microbiota	Collective ecosystem of microorganisms that inhabit the human body	4
Periodontitis	Periodontal disease (PD), or periodontitis, is a chronic inflammatory disease of the periodontium – the tissues that surround and support the teeth	5
Caries	Demineralsation and destruction of enamel and tooth structure by acid-producing caries-associated bacteria (i.e. <i>Streptococcus mutans</i> )	2
Gingivitis	Inflammation, irritation and swelling of the gingiva – the part of the gum that surrounds the base of teeth	6
Orange complex	Microbial community associated with periodontitis. Bacteria in the orange complex are recognised as ‘bridge species’ that bridge non-pathogenic bacteria to pathogenic species through cell-cell interactions	7
Red complex	Microbial community associated with periodontitis. Red complex bacteria include <i>Porphyromonas gingivalis</i> , <i>Treponema denticola</i> , and <i>Tannerella forsythia</i> and are recognised as the most important pathogens in periodontal disease	7

## BACKGROUND

### Oral Microbiota

The human oral microbiome database (HOMD) is the first curated description of human associated microbiome and provides comprehensive information on the approximately 1,000 species that inhabit the human oral cavity (HOMD). The oral microbiota is a complex ecosystem comprised of bacteria, fungi, archaea, viruses, and protozoa, with bacteria being the most studied<sup>8,9</sup>. Bacteria are found on all oral tissues and the composition varies significantly with age<sup>10</sup>. In a recent study, 16S rRNA analysis detected 619 phyla in the oral microbiota; the six main phyla were *Firmicutes*, *Bacteroidetes*, *Proteobacteria*, *Actinobacteria*, *Spirochaetes*, and *Fusobacteria*, which comprised 96% of the taxa. The remaining phyla, *Euryarchaeota*, *Chlamydia*, *Chloroflexi*, SR1, *Synergistetes*, *Tenericutes*, and TM7, comprised the remaining 4% of the taxa<sup>8</sup>. Bacteria associated with periodontal health include *Streptococcus*, *Granulicatella*, *Neisseria*, *Haemophilus*, *Corynebacterium*, *Rothia*, *Actinomyces*, *Prevotella*, and *Capnocytophaga*<sup>11</sup>.

### Homeostasis

The majority of oral microbiota species exist within a complex multispecies structure termed the oral biofilm (dental plaque). The development of oral biofilms is initiated by adherence of pioneer species to salivary proteins and glycoproteins adsorbed onto tooth enamel. In healthy individuals, the oral biofilm is dominated by commensal bacteria that maintain homeostasis through intricate interactions with host immunity and other microorganisms<sup>12</sup>. However, when equilibrium is compromised (drugs, diet, injury, infection, etc.) and microbial imbalance occurs, pathogenic bacteria may colonise and cause pathologies such as dental caries or periodontitis<sup>13</sup>.

Early colonisers of the mouth are commensal *Streptococci* spp. including *S. mitis*, *S. sanguinis* and *S. gordonii*. These early colonisers are able to bind to the tooth surface and prevent colonisation of other bacteria through production of bacteriocins, H<sub>2</sub>O<sub>2</sub> and alkali production<sup>2</sup>. In the absence of a carbohydrate-rich diet, commensal *Streptococci* remain at high levels within dental plaque and maintain homeostasis. Dominance of commensal *Streptococci* is associated with good oral health<sup>2</sup>.

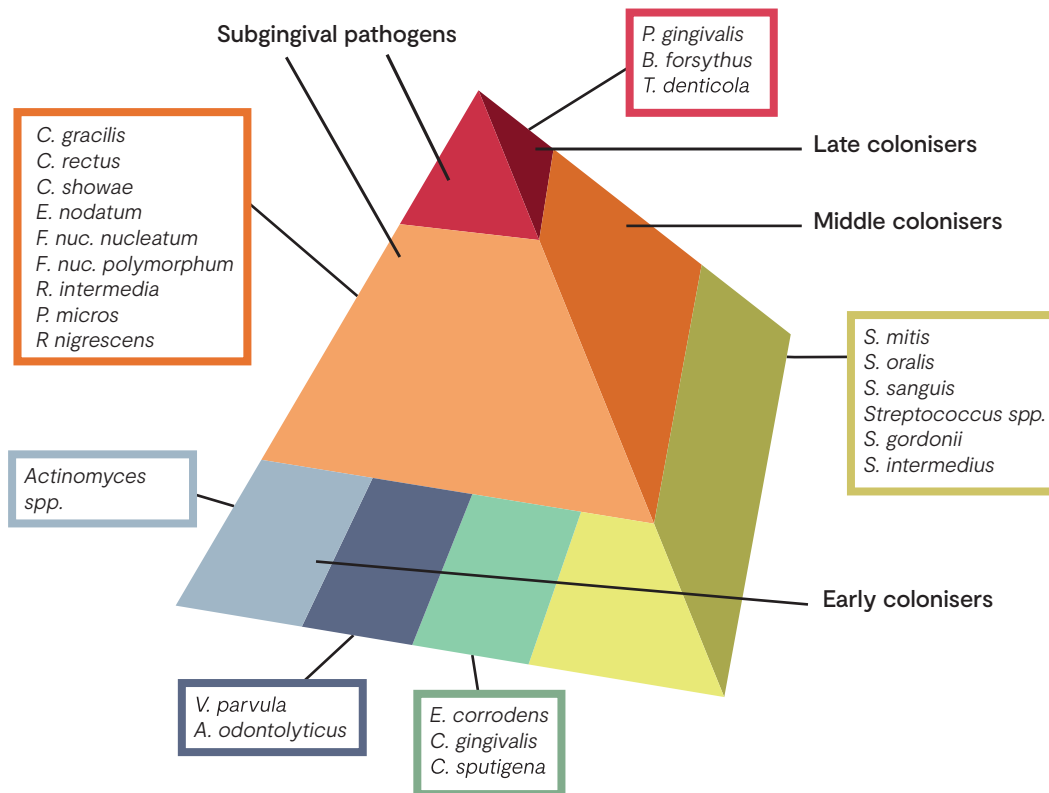
Late colonisers, including Gram-negative (i.e. *Veillonella* spp.) and Gram-positive species (i.e. *Actinomyces* spp.) build up on the initial Streptococcal biofilm via cell-cell interactions. This series of co-aggregation and co-adhesion events leads to the formation of a mature biofilm, a relatively stable community where all species exist in homeostasis<sup>14</sup>. The disruption of homeostasis, termed dysbiosis, has been linked to a multitude of conditions including dental caries, gingivitis, periodontitis and increased risk of oral cancer<sup>15</sup>.

Dysbiosis

Periodontal Disease

Periodontal disease (PD), or periodontitis, is a chronic inflammatory disease of the periodontium, the tissues that surround and support the teeth<sup>5</sup>. It affects 10–15% of adults and is the most common cause of tooth loss worldwide. Periodontal disease has a polymicrobial aetiology within the framework of a complex microbial ecosystem. It is caused by a synergistic and dysbiotic biofilm community, with keystone pathogens such as *Porphyromonas gingivalis* initiating the disruption of tissue homeostasis<sup>16</sup>. The consortium of bacteria most strongly implicated in the pathogenesis of PD is *P. gingivalis*, *T. denticola* and *T. forsythia* (red complex), all three of which, are routinely found in subgingival plaque in patients with chronic periodontitis<sup>7</sup> (Figure 1).

PD is characterised by the destruction of the periodontal ligament, connective tissue and alveolar bone as a result of chronic immune and inflammatory responses. Inflammation in periodontitis is predominantly mediated by IL-1, IL-8, TNF- $\alpha$ , prostaglandins and matrix metalloproteinases (MMPs)<sup>17</sup>. These mediators are proposed to affect functions and activities of leukocytes, osteoblasts and osteoclasts, promoting tissue remodelling locally and systemically<sup>18,19</sup>. Susceptibility to periodontal disease is influenced by host genotype, stress, diet and associated behaviour, including smoking<sup>20</sup>.



**Figure 1:** Periodontitis-associated bacteria grow as synergistic and multispecies communities comprised of different functional complexes dependent on pathogenicity. Figure adapted from Haffajee and Socransky (1994).

Early colonisers include members of the yellow, green, and purple complexes (oral health). The orange complex bacteria generally appear after the early colonisers and include many putative periodontal pathogens, such as *Fusobacterium nucleatum*. The orange complex includes bacteria which, as “bridge species”, form a link between the early colonisers and the highly pathogenic bacteria of the red complex. The pathogenic potential of orange complex bacteria is significantly increased due to the production of various toxins and enzymes. Occurrence of bacteria of the red complex is characteristic of the final colonisation phase, which culminates in the development of a structured, stable bacterial community (climax community) and includes putative periodontal pathogens. Colonisation with these bacteria, which are significantly involved in the destruction of the periodontium, builds on the presence of the less pathogenic species mentioned above. Their ability to penetrate tissue also makes treatment difficult<sup>21</sup>.

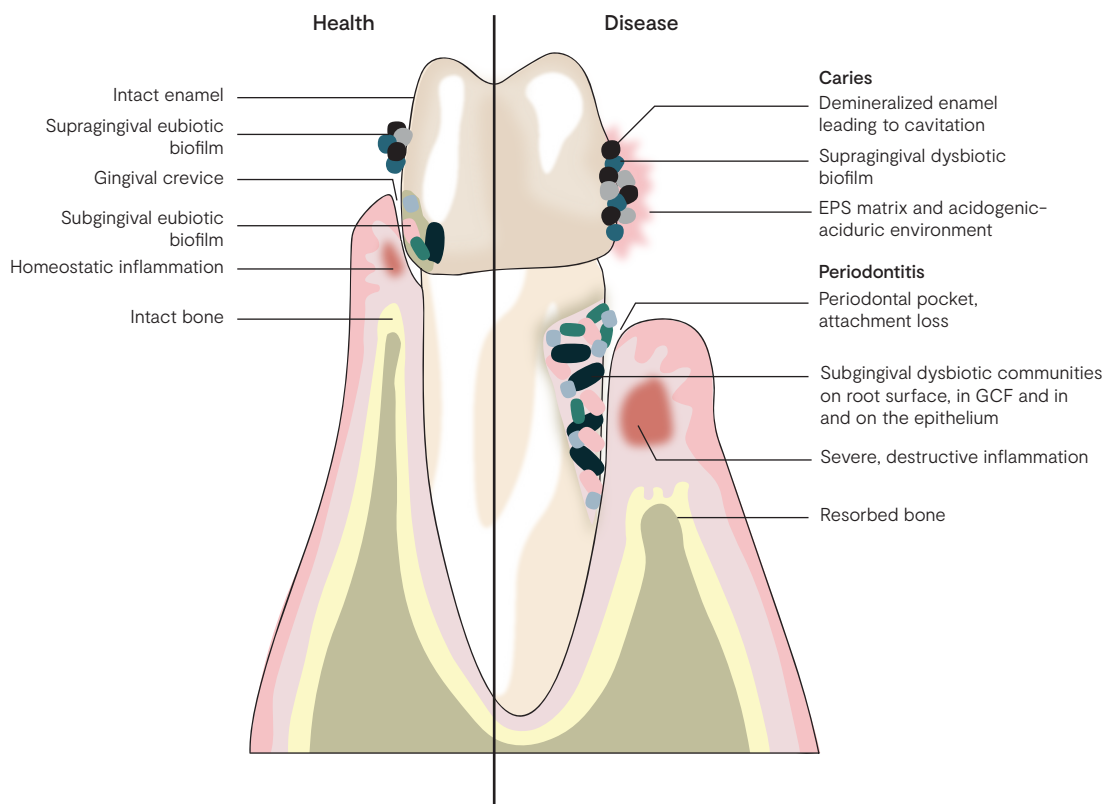
**Caries**

Caries is the most common chronic childhood infection and can lead to progressive destruction of dental hard tissue. Risk factors for caries include the frequent consumption of fermentable dietary carbohydrates (especially sucrose) and/or reduced saliva flow. The increased intake of carbohydrates, alongside poor oral hygiene, leads to increased production of extracellular glucan matrix by oral *Streptococci*. This thickening matrix encapsulates bacteria and creates acidic microenvironments via increased carbohydrate fermentation, that are not readily permeated by saliva. This increasingly acidic environment is not tolerated by commensal *Streptococci* spp. and favours growth of acid-tolerant, caries-associated species including *Streptococcus mutans*, *Veillonella* spp. and *Lactobacillus* species. Species of *Lactobacillus* associated with childhood dental caries include *L. rhamnosus*, *L. casei*, and *L. pseudoplantarum*<sup>22</sup>. Species associated with adulthood caries include *L. casei*, *L. fermentum* and *L. rhamnose*<sup>23,24</sup>. If excessive carbohydrate consumption and poor oral hygiene is prolonged, this results in development of a thick glucan matrix, extremely acidic microenvironments and unrepairable damage to tooth enamel and the onset of clinical disease<sup>2</sup>.

**Oral Cancer**

Oral cancer is one of the ten most prevalent cancers in the world, with more than 90% of mouth neoplasms being oral squamous cell carcinoma (OSCC), originating from the oral mucosa. The pathogenesis of OSCC is attributed mainly to smoking, high alcohol intake and smokeless edible tobacco products. However, other risk factors include viral infections, infection with *Candida* species, periodontitis, poor oral hygiene and chronic bacterial infections and inflammation<sup>25</sup>. Human papillomavirus virus plays a significant role in the development of oral cancer. HPV-16 genotype is the most common genotype to persist in the oral mucosa and is the most common genotype isolated from oral squamous cell carcinomas<sup>26</sup>.

There is an increasing body of evidence supporting a role for *S. intermedius* in oral cancer. In a recent study using biopsies of patients with oral squamous cell carcinoma, *S. intermedius* was detected in 70% of both non-tumour and tumour sites<sup>25</sup>. *T. denticola* has also been detected in oral and gastrointestinal tumour samples<sup>27</sup>.



**Figure 2:** Development of disease in the oral cavity. Oral health involves colonisation of the enamel and gingival crevice by commensal *Streptococci* and bacteria. Dysbiosis involving increased intake of carbohydrates, infection or injury facilitate

colonisation of caries-associated and acid-producing bacteria including *Streptococcus mutans*, causing destruction of the enamel. Dysbiosis may also lead to microbial invasion of the mucosa and further development of subgingival biofilm, involving colonisation by “red complex” pathogens including *P. gingivalis* and *T. denticola*, leading to inflammation and destruction of tooth supporting tissues. Figure adapted from Lamont *et al.* (2018)<sup>28</sup>.

## SYSTEMIC DISORDERS

There is a growing body of research to support a role for PD pathogens as contributing factors for numerous systemic diseases, including cardiovascular disease (CVD), diabetes, adverse pregnancy outcomes, rheumatoid arthritis (RA) and Alzheimer's disease (AD)<sup>29-33</sup>. The proposed aetiology involves direct and indirect effects of oral bacteria on host immune responses, inflammation and therefore development of systemic disorders.

1. Direct mechanism: In chronic periodontitis, periodontal pockets become ulcerated providing direct entry points for oral bacteria into the systemic circulation. Circulating bacteria (i.e. *P. gingivalis*; *T. forsythia*) may then induce direct effects on organs and tissues.
2. Indirect mechanism: Chronic inflammation has been implicated in a multitude of disorders including diabetes, obesity and autoimmune disorders. Chronic inflammation associated with chronic periodontitis may represent a source of inflammation, able to induce and exacerbate systemic disorders.

### Pregnancy

*F. nucleatum* is the most prevalent oral species implicated in adverse pregnancy outcomes (i.e. miscarriage, preterm labour)<sup>34</sup>. It has been detected in a wide variety of placental and foetal tissues including the amniotic fluid and foetal membranes<sup>35</sup>. It is proposed that *F. nucleatum* translocates from the maternal oral cavity to the intrauterine cavity via hematogenous transmission<sup>36</sup>. *C. rectus* is also associated with adverse pregnancy outcomes, including preterm delivery<sup>37</sup>.

### Cardiovascular

*S. mutans* is associated with bacteraemia and infective endocarditis (IE)<sup>38</sup>. These oral bacteria can disseminate into the bloodstream via professional dental treatment and tooth brushing. Indeed, in a review of 848 cases of IE, *Streptococcus* spp. were the most common microorganisms isolated (49.5%)<sup>39</sup>, and *S. mutans* is recovered from 15% of patients with streptococcal valvular disease<sup>40</sup>. The ability of *S. mutans* to produce amyloid as part of the oral biofilm has also implicated this bacterium in dementia<sup>41</sup>. Furthermore, collagen binding surface Cnm protein, expressed on cnm-positive *S. mutans*, has been associated with cerebral microbleeds (CMBs) – an important risk factor for strokes and dementia. In a screen of 279 community residents, cnm-positive *S. mutans* was detected more frequently in patients with CMBs and the risk of CMBs was higher in the group with cnm-positive *S. mutans*<sup>42</sup>. Additionally, systemic infection by *Actinobacillus actinomycetemcomitans* has been associated with atherosclerosis<sup>43</sup>, brain abscesses<sup>53</sup>, and endocarditis<sup>44</sup>.

### Neurodegeneration

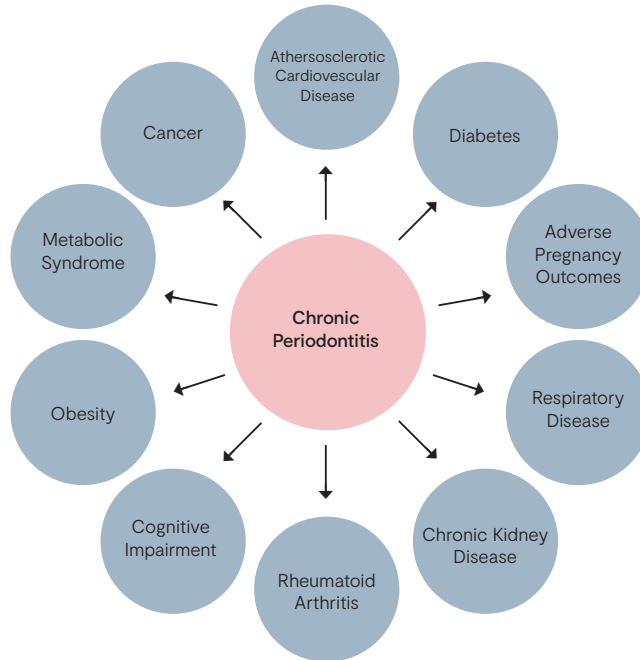
Periodontal pathogens, *P. gingivalis* and *T. denticola* are also associated with the development of Alzheimer's disease (AD)<sup>45</sup>. LPS of *P. gingivalis* was detected in the brain tissue of AD patients<sup>46</sup>. In addition, in a recent study using a mouse model for periodontitis, after infection a significant number of *P. gingivalis* genomic DNA was detected in the brain<sup>47</sup>.

### Arthritis

*P. gingivalis* has also been implicated in the pathogenesis of rheumatoid arthritis (RA). *P. gingivalis* expresses peptidyl arginine deiminase (PAD), which mediates citrullination of peptides. Systemic circulation of citrullinated peptides is a key risk factor for development of RA-associated inflammation<sup>95</sup>. Furthermore, significantly higher anti-*P. gingivalis* antibodies have been detected in patients with RA, compared to systemically and periodontally healthy controls<sup>48</sup>.

**IBD**

*Fusobacterium nucleatum* has been linked to colorectal cancer (CRC). It is found in high amounts in CRC carcinomas, adenomas, in stools of patients with carcinoma and in rectal swabs of CRC patients<sup>49,50</sup>. This bacterium has also been detected in colonic biopsies of IBD patients<sup>51</sup>, and strains isolated from inflamed tissues of IBD patients are more virulent than those from healthy individuals<sup>52</sup>.



**Figure 3:** Associations between periodontitis and systemic disease. Figure adapted from Winning and Linden (2015)<sup>53</sup>.

**METHODOLOGY**

**qRT-PCR FOR QUANTIFICATION OF ORAL MICROBIOTA SPECIES**

The Phylobioscience Oral EcologiX™ profile utilises real-time quantitative PCR (qRT-PCR) for analysis of oral microbiota populations. qRT-PCR is used to quantify the number of copies of a gene of interest in a community sample. Real time qPCR reactions are performed using Taqman technology.

**ELISA ASSAY FOR MEASUREMENT OF PRO-INFLAMMATORY MARKERS**

BIOMARKER	TYPE	CLASSIFICATION
HOST IMMUNE MARKERS		
IL-1β	Pro-inflammatory cytokine	N/A
MICROBIOTA		
Commensal fungi		
<i>Candida albicans</i>	Fungi	N/A

**Bacteria**

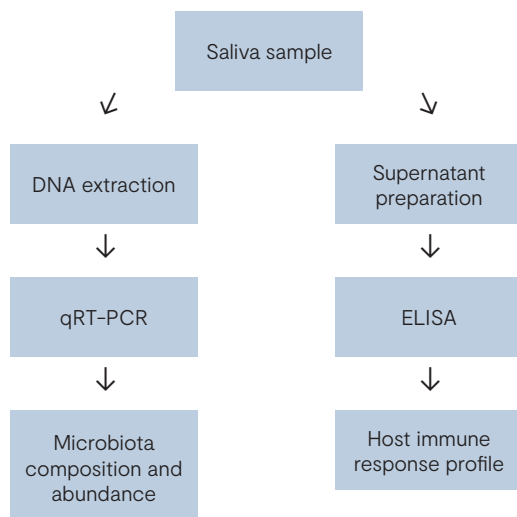
<i>Propionibacterium acidifaciens</i>	Bacteria	Gram-positive
<i>Lactobacillus spp.</i>	Bacteria	Gram-positive
<i>Streptococcus mutans</i>	Bacteria	Gram-positive
<i>Eubacterium nodatum</i>	Bacteria	Gram-positive
<i>Peptostreptococcus anaerobius</i>	Bacteria	Gram-positive
<i>Parvimonas micra</i>	Bacteria	Gram-positive
<i>Porphyromonas gingivalis</i>	Bacteria	Gram-positive
<i>Fusobacterium nucleatum</i>	Bacteria	Gram-negative
<i>Aggregatibacter actinomycetemcomitans</i>	Bacteria	Gram-negative
<i>Treponema denticola</i>	Bacteria	Gram-negative
<i>Actinobacillus actinomycetemcomitans</i>	Bacteria	Gram-negative
<i>Campylobacter rectus</i>	Bacteria	Gram-negative
<i>Tannerella forsythia</i>	Bacteria	Gram-negative
<i>Prevotella intermedia</i>	Bacteria	Gram-negative
<i>Prevotella nigrescens</i>	Bacteria	Gram-negative

**Viral pathogens**

HSV-1	Virus	N/A
HSV-2	Virus	N/A
HPV-16_E6	Virus	N/A
HPV-16_E7	Virus	N/A

Table 1: Oral EcologiX™ biomarkers

**METHODOLOGY FLOWCHART**





## INTERPRETATION OF DATA

### Host Biomarkers

IL-1 $\beta$	HEALTHY: <300pg/ml	HIGH: >300pg/ml
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Interleukin 1- $\beta$  (IL-1 $\beta$ ) is a key mediator of the inflammatory response and a master cytokine that regulates induction of other cytokines, including IL-8<sup>54</sup>. Produced by innate immune cells, it is crucial for host responses against infection and injury. IL-1 $\beta$  is synthesised in response to inflammatory stimuli from pathogens, stress conditions, and other danger signals. IL-1 $\beta$  is a major mediator of inflammation in periodontitis<sup>55</sup>.

### Microbiota Profiles

#### Fungi

<i>Candida albicans</i>	COMMENSAL FUNGI	Overgrowth associated with oral candidiasis
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*Candida albicans* is a fungal species of the commensal oral microbiota and occurs in the oral cavity of 30–100% of healthy individuals<sup>9,63</sup>. The relative abundance of *Candida* species in the oral cavity is influenced by geographic location, health status and diagnosis and exposure to antifungal drugs<sup>64,65</sup>. Additionally, the rate of carriage increases with age; *C. albicans* are recovered from 60% of dental patient's mouths over the age of 60 years<sup>66</sup>.

*C. albicans* is an opportunistic pathogen and can cause oral candidiasis in immunocompromised hosts. Oral candidiasis is associated with changes in oral fungal biodiversity, increased nutrient availability, overgrowth of *Candida* species and transition from yeast to hyphal growth mode, termed phenotypic switching<sup>67</sup>. Several studies have reported increased prevalence of subgingival *C. albicans* in chronic periodontitis patients compared to healthy individuals<sup>68,69</sup>.

#### Gram-positive bacteria

<i>Lactobacillus</i> spp.	COMMENSAL BACTERIA	Associated with dental caries
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*Lactobacillus* spp. are Gram-positive, rod shaped bacteria that comprise a normal member of the human microbiota, inhabiting multiple environments including the gut, genital tract and oral cavity. Lactobacilli were the first microorganisms implicated in dental caries<sup>73</sup>, and subsequent studies reported a strong correlation between *Lactobacillus* counts and dental caries<sup>74,75</sup>. Lactobacilli have been reported to occur in high numbers in both superficial and deep caries<sup>76</sup>.

Previous studies using 16S rRNA sequencing has reported dominant species in child and adult caries to include *L. fermentum*, *L. rhamnosus*, *L. gasseri*, *L. casei* and *L. salivarius*. Less common species include *L. mucosae*, *L. crispatus*, and *L. ultunesis*<sup>77–79</sup>. Low levels of Lactobacilli have been reported in dental plaque from caries-free children<sup>80</sup>.

<i>Streptococcus mutans</i>	COMMENSAL BACTERIA	Associated with dental caries and systemic diseases
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The key habitats for *S. mutans* are the mouth, pharynx and intestine. *S. mutans* has a key role in the aetiology of dental caries as they can adhere to the enamel salivary pellicle and to other plaque bacteria<sup>83</sup>. Appearance of *S. mutans* in the tooth cavity is typically followed by caries after approximately 6–24 months<sup>84</sup>. Acidogenic *S. mutans* is able to form the structural biofilm component, exopolysaccharide (EPS), in the presence of sucrose, fructose and glucose.

In addition to caries, *S. mutans* is linked to a wide range of systemic diseases and disorders including bacteraemia, infective endocarditis and dementia<sup>38–42</sup>.

<i>Eubacterium nodatum</i>	PUTATIVE PATHOGEN	Isolated from patients with severe periodontitis
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*Eubacterium nodatum* are Gram-positive, anaerobic bacteria isolated from patients with moderate and severe adult periodontitis. In a recent study characterising microbiota profiles associated with peri-implantitis, *E. nodatum* was shown to be among the most prevalent species detected<sup>85</sup>. A strong association was shown in periodontitis with *E. nodatum* and *T. denticola* in the presence, or absence, of high levels of core periodontitis pathogens<sup>86</sup>.

<i>Parvimonas micra</i>	COMMENSAL BACTERIA	Associated with periodontitis and systemic diseases
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Originally classified as *Peptostreptococcus micros*, the Gram-positive anaerobic cocci currently classified as *Parvimonas micra*, is a normal member of the gut and oral microbiota. *P. micra* has been implicated in periodontal disease and infections outside the oral cavity<sup>87</sup>. *P. micra* shows significantly higher prevalence in patients with periodontitis<sup>88</sup>. There is a growing body of research that supports the role of *P. micra* as the single causative agent of spondylodiscitis (rare spine infection)<sup>87,89,90</sup>. In rare cases, *P. micra* has been identified as the single causative agent in infectious endocarditis<sup>91</sup>.

<i>Peptostreptococcus anaerobius</i>	COMMENSAL BACTERIA	Associated with periodontitis
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Gram-positive anaerobic cocci frequently cultured from the mouth, upper respiratory tract, skin and soft tissues, bones and joints, GI tract and genitourinary tracts<sup>92</sup>. *P. anaerobius* is associated with periodontitis and has been detected in periodontal disease-associated biofilm<sup>4</sup>. It has been isolated from patients with gingivitis, chronic periodontitis and aggressive periodontitis<sup>4</sup>.

#### Gram-negative bacteria

<i>Fusobacterium nucleatum</i>	COMMENSAL BACTERIA	Associated with periodontitis and systemic disorders
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*Fusobacterium nucleatum* is a Gram-negative obligate anaerobe bacterium that inhabits the oral cavity. *F. nucleatum* is one of the most abundant species in the oral cavity in both diseased and healthy individuals<sup>58</sup>. It is implicated in different types of periodontal diseases including gingivitis, chronic periodontitis, localised aggressive periodontitis and generalised aggressive periodontitis<sup>58-60</sup>. Prevalence of this bacterium increases with the severity of disease, inflammation and pocket depth<sup>5,61</sup>.

*F. nucleatum* is the most prevalent oral species implicated in adverse pregnancy outcomes (i.e. miscarriage, preterm labour)<sup>34</sup>. *F. nucleatum* has also been linked to colorectal cancer<sup>49,50</sup> and IBD<sup>51,52</sup>. *F. nucleatum* is a potent stimulator of inflammatory cytokines, IL-6, IL-8, and TNF- $\alpha$ <sup>62</sup>.

<i>Porphyromonas gingivalis</i>	PATHOGEN	Major periodontitis pathogen
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*Porphyromonas gingivalis* is a Gram-negative anaerobe and is one of the major pathogens associated with chronic periodontitis. Recent work indicates that *P. gingivalis* functions as a keystone organism in periodontitis, i.e. it is an organism able to hold an entire arch/community together. Through expression of a wide range of virulence factors, *P. gingivalis* induces inflammation, evades the host immune response and stimulates bone reabsorption. Local inflammation induces dysbiosis and imbalance of the plaque (biofilm) microbiota<sup>93</sup>. This pathophysiology is recognised as a key initiating event for periodontitis<sup>94</sup>.

*P. gingivalis*, *T. forsythia* and *T. denticola* are categorised as the 'red complex' because they are the most frequently isolated species from chronic lesions in chronic periodontitis<sup>7</sup>. There is also a growing body of evidence linking this pathogen to Alzheimer's Disease, and rheumatoid arthritis<sup>47,48,95</sup>. An association between elevated *P. gingivalis* serum IgG antibodies and increased risk of mortality from oriodigestive cancer has also been reported<sup>96</sup>.

<i>Aggregatibacter actinomycetemcomitans</i>	COMMENSAL BACTERIA	Associated with periodontitis and systemic diseases
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Gram-negative oral pathobiont that causes localised aggressive periodontitis (LAP), characterised by rapid destruction of tooth supporting tissues<sup>97</sup>. LAP has a prevalence of approximately 0.5%, but is 15 times more prevalent in African-American adolescents, compared to the general population<sup>98</sup>. *A. actinomycetemcomitans* is also associated with a range of systemic diseases, including infections of the heart<sup>99</sup>, urinary tract<sup>100</sup> and brain<sup>97,101</sup>.

<i>Treponema denticola</i>	PATHOGEN	Major periodontitis pathogen
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Gram-negative spirochete bacteria frequently isolated from human periodontal lesions. Its role in pathogenesis and progression of periodontitis has been widely reported<sup>102</sup>. *P. gingivalis*, *T. forsythia* and *T. denticola* are categorised as the 'red complex' because they are the most frequently isolated species from chronic lesions in chronic periodontitis<sup>7</sup>. *T. denticola* possess several virulence factors that are implicated in periodontal pathogenesis<sup>103,104</sup>. *T. denticola* is associated with Alzheimer's disease<sup>45</sup> and oral and gastrointestinal cancer development<sup>27,105</sup>.

<i>Actinobacillus actinomycetemcomitans</i>	PATHOGEN	Associated with periodontitis and systemic diseases
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*A. actinomycetemcomitans* is commonly found in the oral microbiota; however, it is recognised as a periodontal pathogen involved in the development of aggressive periodontitis<sup>106,107</sup>. Carriage rates of *A. actinomycetemcomitans* can be up to 20% of the population<sup>108</sup>. Seven serotypes exist, with large variations in their surface antigens and therefore, virulence and pathogenicity<sup>109</sup>. It has been proposed that *A. actinomycetemcomitans* colonises the oral mucosa and translocates to the gingival margin, inducing host immune responses and local inflammation, subsequently leading to degradation of tooth supporting tissues<sup>110</sup>. Systemic infection by this bacterium has been associated with atherosclerosis, brain abscesses<sup>111</sup> and endocarditis<sup>44</sup>.

<i>Campylobacter rectus</i>	PATHOGEN	Periodontitis associated pathogen
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Gram-negative bacteria recognised as a major pathogen in periodontal disease. Together with other oral pathogens, *C. rectus* is implicated in adult periodontitis and rapidly advancing periodontitis<sup>112</sup>. *C. rectus* is often detected in larger numbers in subgingival pockets<sup>113</sup>. It is also associated with adverse pregnancy outcomes, including preterm delivery<sup>37</sup>.

<i>Tannerella forsythia</i>	PATHOGEN	Major periodontitis pathogen
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Alongside *Porphyromonas gingivalis*, *Tannerella forsythia* is widely regarded as a major pathogen. *T. forsythia* is an anaerobic, Gram-negative bacterium isolated from the gingival sulci and periodontal pockets of patients with periodontitis. Three pathogens (*T. forsythia*, *P. gingivalis*, and *T. denticola*), known collectively as the "red complex", are strongly associated with the pathogenesis and progression of destructive forms of periodontitis. The presence of oral *T. forsythia* is associated with an increased risk of oesophageal cancer<sup>114</sup>.

<i>Prevotella spp.</i> ( <i>P. intermedia</i> , <i>P. nigrescens</i> )	COMMENSAL BACTERIA	Associated with periodontitis
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Gram-negative bacteria implicated in various forms of human periodontal disease, including chronic periodontitis<sup>115</sup>, early-onset periodontitis<sup>6</sup> and pregnancy gingivitis. *P. intermedia* and *P. nigrescens*, are members of the "orange complex", and are among the most frequently encountered species in subgingival plaque<sup>116</sup>. *P. intermedia* is a periodontitis-associated member of the subgingival microbiome<sup>117</sup>. Uzel et al. (2011) found that members of the green and orange complexes, such as *Prevotella intermedia*, increased much faster in periodontitis subjects than in periodontally healthy subjects<sup>118</sup>.

**Viral Pathogens**

<i>HSV-1</i>	PATHOGEN	Causative agent of oral herpes
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Herpes simplex virus 1 (HSV-1) is the causative agent of oral herpes. HSV-1 is a dsDNA virus that is prevalent in approximately 90% of the population. HSV-1 infection is often asymptomatic. Infection with HSV-1 may manifest as herpes lesions on the lips, gums and mouth. HSV-1 can also cause acute herpetic gingivostomatitis, a condition that results in ulcers of the mucous membranes inside the mouth. HSV-1 can be transferred via oral, vaginal and anal sex and through skin contact<sup>19</sup>.

<i>HPV-16_E6 &amp; E7</i>	PATHOGEN	Associated with oral cancer
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Human papillomaviruses (HPV) are a large group of viruses, comprised of over 205 genotypes. HPV genotypes belonging to the *Alphapapillomavirus* group are of most clinical relevance as this group contains most of the mucosal HPVs, associated with cancer. HPV typically infects the basal cells of the epithelium<sup>20</sup> and can be transmitted through sexual activity and horizontal transmission and vertical transmission from mother to new born. HPV infection may be asymptomatic; a study of 4,000 subjects detected HPV in approximately 5% of individuals<sup>21</sup>. The HPV-16 genotype is the most common genotype to persist in the oral mucosa and is the most common genotype isolated from oral squamous cell carcinomas<sup>26</sup>.

**ORAL BIOMARKERS**

Condition	ASSOCIATED WITH															
	Oral Disease	Anxiety	Depression	CRC	IBD	IBS	CFS	Diabetes	Metabolic Syndrome	Bacterial Endotoxemia (LPS)	Atherosclerotic Cardiovascular Disease	Obesity	Adverse pregnancy outcomes	Cognitive	Oral cancer	Rheumatoid Arthritis
Periodontitis	x	x	x		x			x	x	x	x		x		x	
Caries	x	x	x		x					x	x					
Oral Dysbiosis	x			x		x										

KEY	DATA REFERENCES
x	Shows association with clinical condition. x does not represent causation.
	Delgado-Angulo, 2015
	Sundararajan, 2015
	Flemer, 2018
	Chandan, 2017
	Winning, 2015
	Fourie, 2016
	Wang, 2018
	Pussinen, 2007
	Lin, 2016
	Ye et al, 2016

## SUMMARY

The Phylobioscience Oral EcologiX™ Oral Health and Microbiome Profile, is an innovative tool for analysis of the oral microbiota composition and host immune responses. Phylobioscience recommend the profile is used on an annual basis to monitor changes in oral health. The oral health and microbiome profile may also be used as a diagnostic tool in symptomatic patients.

For effective analysis of results, the interpretive guide should be used in combination with Invivo Healthcare's Oral Microbiome Clinical Considerations and the patient's symptomatology and results.

The Oral Microbiome Clinical Considerations provides further information on lifestyle and environmental factors that may influence the oral microbiota. The Oral Microbiome Clinical Considerations also provides recommendations for treatment and management plans for results indicative of dysbiosis, inflammation, and infection.

If you have any queries on patient results or clinical considerations, please contact the Clinical Education team at [support@invivohealthcare.com](mailto:support@invivohealthcare.com)

## RECOMMENDED READING

### The oral microbiota: dynamic communities and host interactions

Lamont, R.J., Koo, H. and Hajishengallis, G., 2018.

*Nature Reviews Microbiology*, p.1.

### The human oral microbiome

Dewhirst, F.E., Chen, T., Izard, J., Paster, B.J., Tanner, A.C., Yu, W.H., Lakshmanan, A. and Wade, W.G., 2010.

*Journal of bacteriology*, 192(19), pp.5002–5017.

### Periodontal-disease-associated biofilm: A reservoir for pathogens of medical importance

Colombo, A.P.V., Magalhães, C.B., Hartenbach, F.A.R.R., do Souto, R.M. and da Silva-Boghossian, C.M., 2016.

*Microbial pathogenesis*, 94, pp.27–34.

## REFERENCES

1. Dewhirst, F. E. et al. The human oral microbiome. *J. Bacteriol.* 192, 5002–17 (2010).
2. Baker, J. L. & Edlund, A. Exploiting the Oral Microbiome to Prevent Tooth Decay: Has Evolution Already Provided the Best Tools? *Front. Microbiol.* 9, 3323 (2019).
3. Klimesova, K., Zakostelska, Z. J. & Tlaskalova-Hogenova, H. Oral bacterial and fungal microbiome impacts colorectal carcinogenesis. *Front. Microbiol.* 9, 1–13 (2018).
4. Vieira Colombo, A. P., Magalhães, C. B., Hartenbach, F. A. R. R., Martins do Souto, R. & Maciel da Silva-Boghossian, C. Periodontal-disease-associated biofilm: A reservoir for pathogens of medical importance. *Microb. Pathog.* 94, 27–34 (2016).
5. Yang, N.-Y., Zhang, Q., Li, J.-L., Yang, S.-H. & Shi, Q. Progression of periodontal inflammation in adolescents is associated with increased number of *Porphyromonas gingivalis*, *Prevotella intermedia*, *Tannerella forsythensis*, and *Fusobacterium nucleatum*. *Int. J. Paediatr. Dent.* 24, 226–233 (2014).
6. White, D. & Mayrand, D. Association of oral *Bacteroides* with gingivitis and adult periodontitis. *J. Periodontol.* 16, 259–265 (1981).
7. Socransky, S. S., Haffajee, A. D., Cugini, M. A., Smith, C. & Kent, R. L. Microbial complexes in subgingival plaque. *J. Clin. Periodontol.* 25, 134–44 (1998).
8. Dewhirst, F. E. et al. The Human Oral Microbiome. *J. Bacteriol.* 192, 5002–5017 (2010).
9. Ghannoum, M. A. et al. Characterization of the Oral Fungal Microbiome (Mycobiome) in Healthy Individuals. *PLoS Pathog.* 6, e1000713 (2010).
10. Xu, Z. & Knight, R. Dietary effects on human gut microbiome diversity. *Br. J. Nutr.* 113, S1–S5 (2015).

11. Segata, N. et al. Composition of the adult digestive tract bacterial microbiome based on seven mouth surfaces, tonsils, throat and stool samples. *Genome Biol.* 13, R42 (2012).
12. Bowen, W. H., Burne, R. A., Wu, H. & Koo, H. Oral Biofilms: Pathogens, Matrix, and Polymicrobial Interactions in Microenvironments. *Trends Microbiol.* 26, 229–242 (2018).
13. Badet, C. & Thebaud, N. B. Ecology of lactobacilli in the oral cavity: a review of literature. *Open Microbiol. J.* 2, 38–48 (2008).
14. Kolenbrander, P. E. Oral Microbial Communities: Biofilms, Interactions, and Genetic Systems. *Annu. Rev. Microbiol.* 54, 413–437 (2000).
15. Philip, N., Suneja, B. & Walsh, L. J. Ecological Approaches to Dental Caries Prevention: Paradigm Shift or Shibboleth? *Caries Res.* 52, 153–165 (2018).
16. Jenkinson, H. F. & Lamont, R. J. Oral microbial communities in sickness and in health. *Trends Microbiol.* 13, 589–595 (2005).
17. Fentoğlu, Ö. et al. Pro-inflammatory cytokine levels in association between periodontal disease and hyperlipidaemia. *J. Clin. Periodontol.* 38, 8–16 (2011).
18. Sorsa, T. et al. Matrix metalloproteinases: Contribution to pathogenesis, diagnosis and treatment of periodontal inflammation. *Ann. Med.* 38, 306–321 (2006).
19. Birkedal-Hansen, H. Role of cytokines and inflammatory mediators in tissue destruction. *J. Periodontal Res.* 28, 500–10 (1993).
20. Hajishengallis, G. Periodontitis: from microbial immune subversion to systemic inflammation. *Nat. Rev. Immunol.* 15, 30–44 (2015).
21. Carrouel, F., Viennot, S., Santamaria, J., Veber, P. & Bourgeois, D. Quantitative Molecular Detection of 19 Major Pathogens in the Interdental Biofilm of Periodontally Healthy Young Adults. *Front. Microbiol.* 7, 840 (2016).
22. Nancy, J. & Dorignac, G. Lactobacilli from the dentin and saliva in children. *J. Clin. Pediatr. Dent.* 16, 107–11 (1992).
23. Botha, S. J. Oral lactobacilli isolated from teenage orthodontic patients. *J. Dent. Assoc. S. Afr.* 48, 177–81 (1993).
24. Ozaki, K. et al. A Quantitative Comparison of Selected Bacteria in Human Carious Dentine by Microscopic Counts. *Caries Res.* 28, 137–145 (1994).
25. Pushalkar, S. et al. Comparison of oral microbiota in tumor and non-tumor tissues of patients with oral squamous cell carcinoma. *BMC Microbiol.* 12, 144 (2012).
26. Syrjänen, S. et al. Human papillomaviruses in oral carcinoma and oral potentially malignant disorders: a systematic review. *Oral Dis.* 17, 58–72 (2011).
27. Nieminen, M. T. et al. *Treponema denticola* chymotrypsin-like proteinase may contribute to orodigestive carcinogenesis through immunomodulation. *Br. J. Cancer* 118, 428–434 (2018).
28. Lamont, R. J., Koo, H. & Hajishengallis, G. The oral microbiota: dynamic communities and host interactions. *Nat. Rev. Microbiol.* 16, 745–759 (2018).
29. Mustapha, I. Z., Debrey, S., Oladubu, M. & Ugarte, R. Markers of Systemic Bacterial Exposure in Periodontal Disease and Cardiovascular Disease Risk: A Systematic Review and Meta-Analysis. *J. Periodontol.* 78, 2289–2302 (2007).
30. Paquette, D. W., Brodala, N. & Nichols, T. C. Cardiovascular disease, inflammation, and periodontal infection. *Periodontol.* 2000 44, 113–126 (2007).
31. Friedewald, V. E. et al. The American Journal of Cardiology and Journal of Periodontology Editors' Consensus: Periodontitis and Atherosclerotic Cardiovascular Disease. *J. Periodontol.* 80, 1021–1032 (2009).
32. Kebschull, M., Demmer, R. T. & Papapanou, P. N. Gum Bug, Leave My Heart Alone!"—Epidemiologic and Mechanistic Evidence Linking Periodontal Infections and Atherosclerosis. *J. Dent. Res.* 89, 879–902 (2010).
33. Zelkha, S. A., Freilich, R. W. & Amar, S. Periodontal innate immune mechanisms relevant to atherosclerosis and obesity. *Periodontol.* 2000 54, 207–221 (2010).
34. Casarin, R. C. V. et al. Subgingival biodiversity in subjects with uncontrolled type-2 diabetes and chronic periodontitis. *J. Periodontal Res.* 48, 30–36 (2013).
35. Rubinstein, M. R. et al. *Fusobacterium nucleatum* Promotes Colorectal Carcinogenesis by Modulating E-Cadherin/-Catenin Signaling via its FadA Adhesin. *Cell Host Microbe* 14, 195–206 (2013).

36. Han, Y. W. Can oral bacteria cause pregnancy complications? *Womens. Health (Lond. Engl)*. 7, 401–4 (2011).
37. Arce, R. M. et al. Characterization of the invasive and inflammatory traits of oral *Campylobacter rectus* in a murine model of fetoplacental growth restriction and in trophoblast cultures. *J. Reprod. Immunol.* 84, 145–53 (2010).
38. Nakano, K., Nomura, R. & Ooshima, T. *Streptococcus mutans* and cardiovascular diseases. *Jpn. Dent. Sci. Rev.* 44, 29–37 (2008).
39. Nakatani, S. et al. Current characteristics of infective endocarditis in Japan: an analysis of 848 cases in 2000 and 2001. *Circ. J.* 67, 901–5 (2003).
40. Bruschi, J. *Microbiology of infective endocarditis and clinical correlates: gram-positive organisms*. CRC PRESS (2007).
41. Oli, M. W. et al. Functional amyloid formation by *Streptococcus mutans*. *Microbiology* 158, 2903–16 (2012).
42. Watanabe, I. et al. Oral Cnm-positive *Streptococcus Mutans* Expressing Collagen Binding Activity is a Risk Factor for Cerebral Microbleeds and Cognitive Impairment. *Sci. Rep.* 6, 38561 (2016).
43. Sparks Stein, P. et al. Serum antibodies to periodontal pathogens are a risk factor for Alzheimer's disease. *Alzheimers. Dement.* 8, 196–203 (2012).
44. Winkelhoff, A. J. & Slots, J. *Actinobacillus actinomycetemcomitans* and *Porphyromonas gingivalis* in nonoral infections. *Periodontol.* 2000 20, 122–135 (1999).
45. Foschi, F. et al. *Treponema denticola* in Disseminating Endodontic Infections. *J. Dent. Res.* 85, 761–765 (2006).
46. Poole, S., Singhrao, S. K., Kesavalu, L., Curtis, M. A. & Crean, S. Determining the Presence of Periodontopathic Virulence Factors in Short-Term Postmortem Alzheimer's Disease Brain Tissue. *J. Alzheimer's Dis.* 36, 665–677 (2013).
47. Poole, S. et al. Active Invasion of *Porphyromonas gingivalis* and Infection-Induced Complement Activation in ApoE<sup>-/-</sup> Mice Brains. *J. Alzheimer's Dis.* 43, 67–80 (2014).
48. Bender, P., Bürgin, W. B., Sculean, A. & Eick, S. Serum antibody levels against *Porphyromonas gingivalis* in patients with and without rheumatoid arthritis – a systematic review and meta-analysis. *Clin. Oral Investig.* 21, 33–42 (2017).
49. Kostic, A. D. et al. Genomic analysis identifies association of *Fusobacterium* with colorectal carcinoma. *Genome Res.* 22, 292–298 (2012).
50. Castellarin, M. et al. *Fusobacterium nucleatum* infection is prevalent in human colorectal carcinoma. *Genome Res.* 22, 299–306 (2012).
51. Tahara, T. et al. *Fusobacterium* Detected in Colonic Biopsy and Clinicopathological Features of Ulcerative Colitis in Japan. *Dig. Dis. Sci.* 60, 205–210 (2015).
52. Strauss, J. et al. Invasive potential of gut mucosa-derived *fusobacterium nucleatum* positively correlates with IBD status of the host. *Inflamm. Bowel Dis.* 17, 1971–1978 (2011).
53. Winning, L. & Linden, G. J. Periodontitis and systemic disease. *BDJ Team* 2, 15163 (2015).
54. Dinarello, C. Biologic basis for interleukin-1 in disease. *Blood* 87, (1996).
55. NOH, M. K. et al. Assessment of IL-6, IL-8 and TNF- $\beta$  levels in the gingival tissue of patients with periodontitis. *Exp. Ther. Med.* 6, 847–851 (2013).
56. Brandtzaeg, P. Mucosal Immunity: Induction, Dissemination, and Effector Functions. *Scand. J. Immunol.* 70, 505–515 (2009).
57. KILIAN, M., REINHOLDT, J., LOMHOLT, H., POULSEN, K. & FRANDSEN, E. V. G. Biological significance of IgA1 proteases in bacterial colonization and pathogenesis: critical evaluation of experimental evidence. *APMIS* 104, 321–338 (1996).
58. Griffen, A. L. et al. Distinct and complex bacterial profiles in human periodontitis and health revealed by 16S pyrosequencing. *ISME J.* 6, 1176–1185 (2012).
59. Kistler, J. O., Booth, V., Bradshaw, D. J. & Wade, W. G. Bacterial Community Development in Experimental Gingivitis. *PLoS One* 8, e71227 (2013).
60. Liu, P. et al. Detection of *Fusobacterium Nucleatum* and *fadA* Adhesin Gene in Patients with Orthodontic Gingivitis and Non-Orthodontic Periodontal Inflammation. *PLoS One* 9, e85280 (2014).
61. Riep, B. et al. Are Putative Periodontal Pathogens Reliable Diagnostic Markers? *J. Clin. Microbiol.* 47, 1705–1711 (2009).
62. Park, S.-R. et al. Diverse Toll-Like Receptors Mediate Cytokine Production by *Fusobacterium nucleatum* and *Aggregatibacter actinomycetemcomitans* in Macrophages. *Infect. Immun.* 82, 1914–1920 (2014).



63. Imabayashi, Y. et al. Molecular analysis of fungal populations in patients with oral candidiasis using next-generation sequencing. *Sci. Rep.* 6, 28110 (2016).
64. Gong, Y.-B. et al. Multilocus sequence typing of *Candida albicans* isolates from the oral cavities of patients undergoing haemodialysis. *Sci. Rep.* 8, 16413 (2018).
65. Pfaller, M. A. & Diekema, D. J. Epidemiology of invasive candidiasis: a persistent public health problem. *Clin. Microbiol. Rev.* 20, 133–63 (2007).
66. Singh, A., Verma, R., Murari, A. & Agrawal, A. Oral candidiasis: An overview. *J. Oral Maxillofac. Pathol.* 18, S81–5 (2014).
67. Slutsky, B., Buffo, J. & Soll, D. R. High-frequency switching of colony morphology in *Candida albicans*. *Science* 230, 666–9 (1985).
68. Canabarro, A. et al. Association of subgingival colonization of *Candida albicans* and other yeasts with severity of chronic periodontitis. *J. Periodontol. Res.* 48, 428–432 (2013).
69. Urzúa, B. et al. Yeast diversity in the oral microbiota of subjects with periodontitis: *Candida albicans* and *Candida dubliniensis* colonize the periodontal pockets. *Med. Mycol.* 46, 783–793 (2008).
70. Downes, J. & Wade, W. G. *Propionibacterium acidifaciens* sp. nov., isolated from the human mouth. *Int. J. Syst. Evol. Microbiol.* 59, 2778–2781 (2009).
71. Aas, J. A. et al. Bacteria of dental caries in primary and permanent teeth in children and young adults. *J. Clin. Microbiol.* 46, 1407–17 (2008).
72. Chen, L. et al. Extensive Description and Comparison of Human Supra-Gingival Microbiome in Root Caries and Health. *PLoS One* 10, e0117064 (2015).
73. OWEN, O. W. A study of bacterial counts (lactobacilli) in saliva related to orthodontic appliances; a preliminary report. *Am. J. Orthod.* 35, 672–8 (1949).
74. Roeters, J., Burgersdijk, R., Truin, G. J. & van 't Hof, M. Dental caries and its determinants in 2-to-5-year-old children. *ASDC J. Dent. Child.* 62, 401–8
75. Russell, J. I., MacFarlane, T. W., Aitchison, T. C., Stephen, K. W. & Burchell, C. K. Caries prevalence and microbiological and salivary caries activity tests in Scottish adolescents. *Community Dent. Oral Epidemiol.* 18, 120–125 (1990).
76. Hahn, C., Jr, W. F., biology, G. M.-A. of oral & 1991, undefined. Microbiological studies of carious dentine from human teeth with irreversible pulpitis. Elsevier
77. Piwat, S., Teanpaisan, R., ... S. T.-M. oral & 2010, undefined. Lactobacillus species and genotypes associated with dental caries in Thai preschool children. Wiley Online Libr.
78. Teanpaisan, R., Chaethong, W., ... S. P.-P. & 2012, undefined. Vertical transmission of mutans streptococci and lactobacillus in Thai families. ingentaconnect.com
79. Yang, R., Argimon, S., Li, Y., ... X. Z.-J. of microbiological & 2010, undefined. Determining the genetic diversity of lactobacilli from the oral cavity. Elsevier
80. Byun, R. et al. Quantitative analysis of diverse Lactobacillus species present in advanced dental caries. *J. Clin. Microbiol.* 42, 3128–36 (2004).
81. Hammes, W. P. & Vogel, R. F. The genus Lactobacillus. in *The Genera of Lactic Acid Bacteria* 19–54 (Springer US, 1995). doi:10.1007/978-1-4615-5817-0\_3
82. Nissen, L., Sgorbati, B., Biavati, B. & Belibasakis, G. N. Lactobacillus salivarius and L. gasseri down-regulate Aggregatibacter actinomycetemcomitans exotoxins expression. *Ann. Microbiol.* 64, 611–617 (2014).
83. Lamont, R. J., Demuth, D. R., Davis, C. A., Malamud, D. & Rosan, B. Salivary-agglutinin-mediated adherence of Streptococcus mutans to early plaque bacteria. *Infect. Immun.* 59, 3446–50 (1991).
84. Balakrishnan, M., Simmonds, R. S. & Tagg, J. R. Dental caries is a preventable infectious disease. *Aust. Dent. J.* 45, 235–245 (2000).

85. Tamura, N., Ochi, M., Miyakawa, H. & Nakazawa, F. Analysis of bacterial flora associated with peri-implantitis using obligate anaerobic culture technique and 16S rDNA gene sequence. *Int. J. Oral Maxillofac. Implants* 28, 1521–9 (2013).
86. Haffajee, A. D., Teles, R. P. & Socransky, S. S. Association of *Eubacterium nodatum* and *Treponema denticola* with human periodontitis lesions. *Oral Microbiol. Immunol.* 21, 269–282 (2006).
87. Uemura, H. et al. *Parvimonas micra* as a causative organism of spondylodiscitis: a report of two cases and a literature review. (2014). doi:10.1016/j.ijid.2014.02.007
88. Rams, T. E., Feik, D., Listgarten, M. A. & Slots, J. *Peptostreptococcus micros* in human periodontitis. *Oral Microbiol. Immunol.* 7, 1–6 (1992).
89. Pilmis, B., Israel, J., Le Monnier, A. & Mizrahi, A. Spondylodiscitis due to anaerobic bacteria about a case of *Parvimonas micra* infection. *Anaerobe* 34, 156–157 (2015).
90. van Duijvenbode, D. C., Kuiper, J. W. P., Holewijn, R. M. & Stadhouder, A. *Parvimonas micra* Spondylodiscitis: A Case Report and Systematic Review of the Literature. *J. Orthop. case reports* 8, 67–71 (2018).
91. Gomez, C. A. et al. First case of infectious endocarditis caused by *Parvimonas micra*. *Anaerobe* 36, 53–55 (2015).
92. Könönen, E., Bryk, A., Niemi, P. & Kanervo–Nordström, A. Antimicrobial susceptibilities of *Peptostreptococcus anaerobius* and the newly described *Peptostreptococcus stomatis* isolated from various human sources. *Antimicrob. Agents Chemother.* 51, 2205–7 (2007).
93. Zenobia, C. & Hajishengallis, G. *Porphyromonas gingivalis* virulence factors involved in subversion of leukocytes and microbial dysbiosis. *Virulence* 6, 236–243 (2015).
94. Darveau, R. P., Hajishengallis, G. & Curtis, M. A. *Porphyromonas gingivalis* as a Potential Community Activist for Disease. *J. Dent. Res.* 91, 816 (2012).
95. Lundberg, K. et al. Antibodies to citrullinated  $\beta$ -enolase peptide 1 are specific for rheumatoid arthritis and cross-react with bacterial enolase. *Arthritis Rheum.* 58, 3009–3019 (2008).
96. Ahn, J., Segers, S. & Hayes, R. B. Periodontal disease, *Porphyromonas gingivalis* serum antibody levels and orodigestive cancer mortality. *Carcinogenesis* 33, 1055–1058 (2012).
97. Fine, D. H., Patil, A. G. & Velusamy, S. K. *Aggregatibacter actinomycetemcomitans* (Aa) Under the Radar: Myths and Misunderstandings of Aa and Its Role in Aggressive Periodontitis. *Front. Immunol.* 10, 728 (2019).
98. Zambon, J. J. *Actinobacillus actinomycetemcomitans* in human periodontal disease. *J. Clin. Periodontol.* 12, 1–20 (1985).
99. Yuan, A. et al. *Actinobacillus Actinomycetemcomitans* Pneumonia with Chest Wall Involvement and Rib Destruction. *Chest* 101, 1450–1452 (1992).
100. Townsend, T., JAMA, J. G.- & 1969, undefined. Urinary tract infection due to *Actinobacillus actinomycetemcomitans*. jamanetwork.com
101. Martin, B. F., Derby, B. M., Budzilovich, G. N. & Ransohoff, J. Brain abscess due to *Actinobacillus actinomycetemcomitans*. *Neurology* 17, 833–7 (1967).
102. Ellen, R. P. & Galimanas, V. B. Spirochetes at the forefront of periodontal infections. *Periodontol.* 2000 38, 13–32 (2005).
103. Fenno, J. C. et al. Cytopathic effects of the major surface protein and the chymotrypsinlike protease of *Treponema denticola*. *Infect. Immun.* 66, 1869–77 (1998).
104. Nussbaum, G., Ben-Adi, S., Genzler, T., Sela, M. & Rosen, G. Involvement of Toll-Like Receptors 2 and 4 in the Innate Immune Response to *Treponema denticola* and Its Outer Sheath Components. *Infect. Immun.* 77, 3939–3947 (2009).
105. Narikiyo, M. et al. Frequent and preferential infection of *Treponema denticola*, *Streptococcus mitis*, and *Streptococcus anginosus* in esophageal cancers. *Cancer Sci.* 95, 569–574 (2004).
106. Genco, R. J. Current View of Risk Factors for Periodontal Diseases. *J. Periodontol.* 67, 1041–1049 (1996).
107. Henderson, B., Ward, J. M. & Ready, D. *Aggregatibacter (Actinobacillus) actinomycetemcomitans*: a triple A\* periodontopathogen? *Periodontol.* 2000 54, 78–105 (2010).
108. Kononen, E. et al. Population-Based Study of Salivary Carriage of Periodontal Pathogens in Adults. *J. Clin. Microbiol.* 45, 2446–2451 (2007).

109. Akrivopoulou, C., Green, I. M., Donos, N., Nair, S. P. & Ready, D. Aggregatibacter actinomycetemcomitans serotype prevalence and antibiotic resistance in a UK population with periodontitis. *J. Glob. Antimicrob. Resist.* 10, 54–58 (2017).
110. Herbert, B. A., Novince, C. M. & Kirkwood, K. L. Aggregatibacter actinomycetemcomitans, a potent immunoregulator of the periodontal host defense system and alveolar bone homeostasis. *Mol. Oral Microbiol.* 31, 207–227 (2016).
111. British Dental Journal. Brain abscess associated with Aggregatibacter actinomycetemcomitans: case report and review of literature. *Br. Dent. J.* 211, 589–589 (2011).
112. Ihara, H. et al. Detection of Campylobacter rectus in periodontitis sites by monoclonal antibodies. *J. Periodontal Res.* 38, 64–72 (2003).
113. Dzink, J. L., Tanner, A. C. R., Haffajee, A. D. & Socransky, S. S. Gram negative species associated with active destructive periodontal lesions. *J. Clin. Periodontol.* 12, 648–659 (1985).
114. Malinowski, B. et al. The role of Tannerella forsythia and Porphyromonas gingivalis in pathogenesis of esophageal cancer. *Infect. Agent. Cancer* 14, 3 (2019).
115. Slots, J. & Genco, R. J. Microbial Pathogenicity Black-pigmented Bacteroides species, Capnocytophaga species, and Actinobacillus actinomycetemcomitans in Human Periodontal Disease: Virulence Factors in Colonization, Survival, and Tissue Destruction. *J. Dent. Res.* 63, 412–421 (1984).
116. Kamma, J. J., Nakou, M., Gmur, R. & Baehni, P. C. Microbiological profile of early onset/aggressive periodontitis patients. *Oral Microbiol. Immunol.* 19, 314–321 (2004).
117. Zheng, H. et al. Subgingival microbiome in patients with healthy and ailing dental implants. *Sci. Rep.* 5, 1–11 (2015).
118. Uzel, N. G. et al. Microbial shifts during dental biofilm re-development in the absence of oral hygiene in periodontal health and disease. *J. Clin. Periodontol.* 38, 612–620 (2011).
119. Arduino, P. & Porter, S. Oral and perioral herpes simplex virus type 1 (HSV-1) infection: review of its management\*. *Oral Dis.* 12, 254–270 (2006).
120. Syrjänen, S. Oral manifestations of human papillomavirus infections. *Eur. J. Oral Sci.* 126, 49 (2018).
121. Kreimer, A., Bhatia, R., ... A. M.-S. transmitted & 2010, undefined. Oral human papillomavirus in healthy individuals: a systematic review of the literature. [journals.lww.com](http://journals.lww.com)